

มาตรฐานผลิตภัณฑ์อุตสาหกรรม

THAI INDUSTRIAL STANDARD

มอก. 2240 เล่ม 1- 2548

ISO/TS 11133-1: 2000

จุลชีววิทยาของอาหารและอาหารสัตว์ – ข้อแนะนำในการเตรียมและผลิตอาหารเลี้ยงเชื้อ

เล่ม 1 ข้อแนะนำทั่วไปในการประกันคุณภาพสำหรับการเตรียมอาหาร เลี้ยงเชื้อในห้องปฏิบัติการ

MICROBIOLOGY OF FOOD AND ANIMAL FEEDING STUFFS – GUIDELINES ON PREPARATION AND PRODUCTION OF CULTURE MEDIA

PART 1 GENERAL GUIDELINES ON QUALITY ASSURANCE FOR THE PREPARATION OF CULTURE MEDIA IN THE LABORATORY

สำนักงานมาตรฐานผลิตภัณฑ์อุตสาหกรรม

มาตรฐานผลิตภัณฑ์อุตสาหกรรม จุลชีววิทยาของอาหารและอาหารสัตว์ – ข้อแนะนำในการเตรียมและผลิตอาหารเลี้ยงเชื้อ

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มอก. 2240 เล่ม 1- 2548

สำนักงานมาตรฐานผลิตภัณฑ์อุตสาหกรรม กระทรวงอุตสาหกรรม ถนนพระรามที่ 6 กรุงเทพฯ 10400 โทรศัพท์ 0 2202 3300

ประกาศในราชกิจจานุเบกษา ฉบับประกาศและงานทั่วไป เล่ม 123 ตอนที่ 12ง วันที่ 9 กุมภาพันธ์ พุทธศักราช 2549

คณะกรรมการวิชาการคณะที่ 957 มาตรฐานอาหารและผลิตภัณฑ์เกษตรด้านจุลชีววิทยา

ประธานกรรมการ

นางเพ็ญศรี รอดมา กรมวิทยาศาสตร์การแพทย์

กรรมการ

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สำนักตรวจสอบคุณภาพสินค้าปศุสัตว์ กรมปศุสัตว์ นายพลายแก้ว ไชยเบญจวงศ์ สถาบันวิจัยวิทยาศาสตร์และเทคโนโลยีแห่งประเทศไทย

นางมาลัย บุญรัตนกรกิจ สถาบันค้นคว้าและพัฒนาผลิตภัณฑ์อาหาร

มหาวิทยาลัยเกษตรศาสตร์

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สำนักตรวจสอบคุณภาพสินค้าปศุสัตว์ กรมปศุสัตว์

นางสาวอัจฉรา วีระยุทธวิไล บริษัท เช้าท์อีสต์เอเชี่ยนลาบอราทอรี่ย์ จำกัด (ซีล)

นางนิตยา พิระภัทรุ่งสุริยา สถาบันอาหาร

นางสาวอรรถยา เกียรติสุนทร กลุ่มตรวจสอบคุณภาพนมและผลิตภัณฑ์นม

สำนักตรวจสอบคุณภาพสินค้าปศุสัตว์ กรมปศุสัตว์

กรรมการและเลขานุการ

นายณฤทธิ์ ฤกษ์ม่วง สำนักงานมาตรฐานผลิตภัณฑ์อุตสาหกรรม

อาหารเลี้ยงเชื้อที่ใช้ในขั้นตอนและวิธีการทดสอบทางจุลชีววิทยาที่มีคุณภาพสม่ำเสมอทำให้ได้ผลการทดสอบที่สามารถ ทวนซ้ำได้เมื่อทดสอบซ้ำ อาหารเลี้ยงเชื้อได้นำไปใช้ในทุกเทคนิคการเพาะเลี้ยงเชื้อและเทคนิคที่เป็นทางเลือกอื่น อีกหลายวิธี อาหารเลี้ยงเชื้อชนิดแห้งหลายชนิดที่มีจำหน่ายทั่วไปและอีกหลายชนิดที่ใช้สำหรับวัตถุประสงค์ ที่เฉพาะเจาะจง และระบุอยู่ในเอกสารวิชาการ นอกจากนี้มีวัตถุประสงค์หลักของห้องปฏิบัติการจุลชีววิทยาอาหาร คือ ทำให้เชื้อจุลินทรีย์หลากหลายชนิด คงสภาพ ฟื้นตัว เจริญเติบโต สามารถตรวจสอบ และ/หรือ นับจำนวนได้ อาหารเลี้ยงเชื้อที่ใช้ต้องมีความเฉพาะเจาะจงต่อตัวอย่างและเชื้อจุลินทรีย์ที่ทำการตรวจสอบ

ทั้งนี้ข้อกำหนดเบื้องต้นสำหรับงานด้านจุลชีววิทยาที่น่าเชื่อถือ อาหารเลี้ยงเชื้อต้องเป็นไปตามข้อกำหนดหรือเกณฑ์ ขั้นต่ำ ข้อกำหนดการทดสอบคุณภาพอาหารเลี้ยงเชื้อควรมีความเพียงพอเพื่อแสดงว่า 1) อาหารเลี้ยงเชื้อแต่ละรุ่น ต้องเป็นที่ยอมรับ 2) อาหารเลี้ยงเชื้อเหมาะสมกับวัตถุประสงค์ของการใช้ 3) อาหารเลี้ยงเชื้อให้ผลการทดสอบ ที่เป็นไปในทางเดียวกันเสมอ

เกณฑ์กำหนดทั้งสาม และการจัดทำระบบเอกสารที่มีความเหมาะสมเป็นส่วนสำคัญของวิธีควบคุมคุณภาพภายใน ซึ่งทำให้การตรวจสอบอาหารเลี้ยงเชื้อมีประสิทธิภาพเป็นผลให้ได้ข้อมูลที่แม่นยำและเที่ยงตรง

คณะกรรมการวิชาการคณะที่ 957 มาตรฐานอาหารและผลิตภัณฑ์เกษตรด้านจุลชีววิทยา รับผิดชอบงานของ คณะอนุกรรมการคณะที่ 9 ภายใต้กรรมการวิชาการคณะที่ 34 ขององค์การระหว่างประเทศว่าด้วยการมาตรฐาน เป็นผู้จัดทำมาตรฐานผลิตภัณฑ์อุตสาหกรรมนี้

มาตรฐานผลิตภัณฑ์อุตสาหกรรมนี้จัดทำขึ้นเพื่อให้ทันกับความต้องการของผู้ใช้ และจักได้แปลเป็นภาษาไทย ในโอกาสอันสมควร หากมีข้อสงสัยโปรดติดต่อสอบถามที่สำนักงานมาตรฐานผลิตภัณฑ์อุตสาหกรรม

คณะกรรมการมาตรฐานผลิตภัณฑ์อุตสาหกรรมได้พิจารณามาตรฐานนี้แล้ว เห็นสมควรเสนอรัฐมนตรีประกาศตาม มาตรา 15 แห่งพระราชบัญญัติมาตรฐานผลิตภัณฑ์อุตสาหกรรม พ.ศ. 2511



ประกาศกระทรวงอุตสาหกรรม ฉบับที่ 3433 (พ.ศ. 2548)

ออกตามความในพระราชบัญญัติมาตรฐานผลิตภัณฑ์อุตสาหกรรม

พ.ศ. 2511

เรื่อง กำหนดมาตรฐานผลิตภัณฑ์อุตสาหกรรม จุลชีววิทยาของอาหารและอาหารสัตว์–ข้อแนะนำในการเตรียมและผลิตอาหารเลี้ยงเชื้อ เล่ม 1 ข้อแนะนำทั่วไปในการประกันคุณภาพสำหรับการเตรียมอาหารเลี้ยงเชื้อในห้องปฏิบัติการ

อาศัยอำนาจตามความในมาตรา 15 แห่งพระราชบัญญัติมาตรฐานผลิตภัณฑ์อุตสาหกรรม พ.ศ. 2511 รัฐมนตรีว่าการกระทรวงอุตสาหกรรมออกประกาศกำหนดมาตรฐานผลิตภัณฑ์อุตสาหกรรม จุลชีววิทยาของอาหาร และอาหารสัตว์–ข้อแนะนำในการเตรียมและผลิตอาหารเลี้ยงเชื้อ เล่ม 1 ข้อแนะนำทั่วไปในการประกันคุณภาพสำหรับ การเตรียมอาหารเลี้ยงเชื้อในห้องปฏิบัติการ มาตรฐานเลขที่ มอก. 2240 เล่ม 1–2548 ไว้ ดังมีรายการละเอียดต่อท้าย ประกาศนี้

ประกาศ ณ วันที่ 31 ตุลาคม พ.ศ. 2548
นายสุริยะ จึงรุ่งเรืองกิจ
รัฐมนตรีว่าการกระทรวงอุตสาหกรรม

เล่ม 1 ข้อแนะนำทั่วไปในการประกันคุณภาพสำหรับการเตรียมอาหาร เลี้ยงเชื้อในห้องปฏิบัติการ

มาตรฐานผลิตภัณฑ์อุตสาหกรรมนี้กำหนดขึ้นโดยรับมาตรฐาน ISO/TS 11133-1:2000 Microbiology of food and animal feeding stuffs- Guidelines on preparation and production of culture media-Part 1: General guidelines on quality assurance for the preparation of culture media in the laboratory มาใช้โดยวิธีพิมพ์ซ้ำในระดับ เหมือนกันทุกประการ

มาตรฐานฉบับนี้ระบุคำศัพท์ทั่วไปที่เกี่ยวข้องกับการประกันคุณภาพของการเตรียมอาหารเลี้ยงเชื้อและข้อกำหนดขั้นต่ำ เฉพาะที่ใช้สำหรับการวิเคราะห์ทางจุลชีววิทยาของผลิตภัณฑ์อาหารหรืออาหารสัตว์

ข้อกำหนดนี้สามารถใช้ได้กับอาหารเลี้ยงเชื้อสามกลุ่มในห้องปฏิบัติการที่มีการเตรียมและ/หรือใช้สำหรับการวิเคราะห์ ทางจุลชีววิทยาได้แก่

- อาหารเลี้ยงเชื้อพร้อมใช้ที่ผลิตขึ้นเพื่อการค้า
- อาหารเลี้ยงเชื้อที่เตรียมจากอาหารเลี้ยงเชื้อชนิดแห้งที่ผลิตขึ้นเพื่อการค้า
- อาหารเลี้ยงเชื้อที่เตรียมขึ้นเองจากส่วนประกอบเฉพาะ

เอกสารฉบับนี้กำหนดคำจำกัดความของการประกันคุณภาพ อาหารเลี้ยงเชื้อ และเชื้อจุลินทรีย์ที่ใช้ในการทดสอบ นอกจากนี้ยังกำหนดข้อปฏิบัติสำหรับการควบคุมคุณภาพของอาหารเลี้ยงเชื้อซึ่งประกอบด้วยการจัดทำ เอกสารสำหรับผู้ผลิตและการทำแบบรายการตรวจสอบโดยห้องปฏิบัติการ การจัดเก็บอาหารเลี้ยงเชื้อในสภาวะ ที่เหมาะสม การเตรียมอาหารเลี้ยงเชื้อ น้ำสำหรับเตรียมอาหารเลี้ยงเชื้อ การฆ่าเชื้อด้วยความร้อนและการกรอง การเตรียมอาหารเลี้ยงเชื้อก่อนใช้งาน การกำจัดอาหารเลี้ยงเชื้อ และขั้นตอนอื่นที่เกี่ยวข้องการควบคุมคุณภาพของ อาหารเลี้ยงเชื้อก่อนใช้งานมีทั้งการควบคุมคุณภาพด้านกายภาพและจุลชีววิทยา

1 Scope

This European prestandard provides the general terminology related to quality assurance of the preparation of culture media and specifies the **minimum** requirements to be used for the microbiological analysis of products intended for human consumption or animal feeding.

These requirements are applicable to three categories of culture media used in laboratories that prepare and/or use culture media for performing microbiological analyses:

- commercially manufactured ready-to-use media;
- media prepared from commercially available dehydrated formulations (either complete e.g. plate count agar or basal media to which supplements are added e.g. Baird-Parker agar);
- media prepared from its individual components.

2 Normative references

This European Prestandard incorporates by dated or undated reference, provisions from other publications. These normative references are cited at the appropriate places in the text and the publications are listed hereafter. For dated references, subsequent amendments to or revisions of any of these publications apply to this European Prestandard only when incorporated in it by amendment or revision. For undated references the latest edition of the publication referred to applies.

EN 1659:1996, In vitro diagnostic systems —Culture media for microbiology —Terms and definitions.

EN 12322:1999, In vitro diagnostic medical devices —Culture media for microbiology —Performance criteria for culture media.

ISO 8402:1994, Quality management and quality assurance —Vocabulary. 1)

3 Terminology

3.1 General

This clause gives the general definitions related to quality assurance and provides different types of terminology related to culture media and to control cultures. Standards cited between brackets indicate that the text given is identical to that cited.

3.2 Terminology of quality assurance

3.2.1

quality assurance

all the planned and systematic activities implemented within the quality system and demonstrated as needed, to provide adequate confidence that an entity will fulfil the requirements for quality

[ISO 8402]

¹⁾ This is under revision and will be combined with ISO 9000-1:1994 to become ISO 9000:2000, *Quality management systems — Fundamentals and vocabulary*.

3.2.2

quality control

operational techniques and activities that are used to fulfil the requirements for quality

[ISO 8402]

3.2.3

internal quality control

a continuous control programme of the laboratory's work prepared by or for them, and based on control analysis together with follow-up and, if necessary, corrective actions

3.2.4

batch of culture media; lot of culture media

fully traceable unit of a medium referring to a defined amount of bulk, semi-finished product or end product, which is consistent in type and quality and which has passed the requirements of production (in-process control) and quality assurance testing, and which has been produced within one defined production period, having been assigned the same lot number

[EN 12322]

3.2.5

performance of culture media

the response of a culture medium to challenge by test organisms under defined conditions

3.3 Terminology of culture media

3.3.1

culture medium

formulation of substances, in liquid, semi-solid or in solid form, which contain natural and/or synthetic constituents intended to support the multiplication, or to preserve the viability, of microorganisms

NOTE When used in connection with compound words, this term is often shortened into "medium" (e.g. enrichment medium).

[EN 1659]

3.3.1 Culture media classified by composition

3.3.2.1

chemically defined culture medium

culture medium consisting only of chemically defined constituents (i.e. of known molecular structure and degree of purity)

[EN 1659]

3.3.2.2

chemically incomplete culture medium

culture medium consisting entirely or partly of natural materials, processed or otherwise, the chemical composition of which is not completely defined

NOTE For the various chemically undefined components used in culture media, ISO/TC 34/SC 9 has specified harmonised designations - see Annex A.

3.3.2 Culture media classified by consistency

3.3.3.1

liquid culture medium

culture medium consisting of an aqueous solution of one or more constituents (e.g. peptone water, nutrient broth)

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NOTE 1 In some cases, solid particles are added to the liquid culture medium.

NOTE 2 Liquid media in tubes, flasks or bottles are commonly called "broth".

[EN 1659]

3.3.3.2

solid culture medium and semi-solid culture medium

liquid culture medium containing solidifying materials (e.g. agar-agar, gelatine, etc.,) in different concentrations

NOTE 1 Due to the world-wide use of culture media solidified with agar-agar, the shortened term "agar" is often used synonymously for solid culture media and therefore in connection with nouns, e.g. "Plate count agar".

NOTE 2 Solid culture media poured into Petri dishes are commonly called "plates". Solid culture media poured into tubes that are kept in slanted positions while the media are solidifying are often called "slants".

[EN 1659]

3.3.3 Culture media classified by intent of use

3.3.4.1

transport medium

culture medium designed to preserve and maintain the viability of microorganisms for the time period between sample collection and laboratory processing of the sample

NOTE Transport media usually contain substances that do not permit multiplication of microorganisms but ensure their preservation (e.g. Stuart's or Amies' Transport medium).

[EN 1659]

3.3.4.2

preservation medium

culture medium designed to preserve and maintain the viability of microorganisms over an extended period, to protect them against the adverse influences which may occur during long-term storage and to allow recovery after this period (e.g. Dorset egg medium)

[EN 1659]

3.3.4.3

resuscitation medium

culture medium enabling stressed and damaged microorganisms to repair and recover their capacity for normal growth without necessarily promoting their multiplication

[EN 1659]

3.3.4.4

enrichment medium

predominantly liquid culture medium which, due to its composition, provides particularly favourable conditions for multiplication of microorganisms

[EN 1659]

3.3.4.4.1

selective enrichment medium

enrichment medium which supports the multiplication of specific microorganisms whilst partially or totally inhibiting the growth of other microorganisms (e.g. Rappaport-Vassiliadis medium)

3.3.4.4.2

non-selective enrichment medium

enrichment medium which supports the growth of most microorganisms (e.g. nutrient broth)

3.3.4.5

isolation medium

solid or semi-solid culture medium which supports the growth of microorganisms

3.3.4.5.1

selective isolation medium

isolation medium which supports growth of specific microorganisms, while inhibiting other microorganisms (e.g. PALCAM agar, MacConkey agar)

[EN 1659]

3.3.4.5.2

non-selective isolation medium

isolation medium which is not devised to selectively inhibit microorganisms (e.g. nutrient agar)

[EN 1659]

3.3.4.6

differential medium

culture medium which permits the testing of one or more physiological/biochemical characteristics of the microorganisms for their identification (e.g. Urea medium, Kligler agar)

NOTE Differential media which can be used as isolation media are referred to as isolation/differential media (e.g. xylose lysine desoxycholate (XLD) agar).

[EN 1659]

3.3.4.7

identification medium

culture medium designed to produce a specific identification reaction which does not require any further confirmatory test

NOTE Identification media which can be used as isolation media are referred to as isolation/identification media.

[EN 1659]

3.3.4.8

media having multiple uses

certain culture media may be assigned to several categories, e.g. Blood agar is a resuscitation medium according to 3.3.4.3. an isolation medium according to 3.3.4.5 and a differential medium according to 3.3.4.6 used for detection of haemolysis

3.3.4 Culture media classified according to preparation method

3.3.5.1

ready-to-use medium

culture medium which is supplied in containers in ready-to-use form (e.g. Petri dishes or tubes or other containers)

3.3.5.2

culture medium prepared from commercially dehydrated formulations

culture medium in dry form which is not ready for immediate use (e.g. powders, granules, lyophilised products). Rehydration will make one of two kinds of medium

- a complete ready-to-use medium;
- an incomplete medium to which labile components are added at the time of use.

3.3.5.3 Culture medium prepared from individual components in the laboratory

3.4 Terminology for test organisms

3.4.1 General

These are microorganisms generally used for quality control and performance testing of culture media. They are defined according to their source as follows.

3.4.2

reference strain

microorganism defined to at least the genus and species level, catalogued and described according to its characteristics and preferably stating its origin

[EN 12322]

3.4.3

reference stocks

a set of separate identical cultures obtained in the laboratory by a single sub-culture from the reference strain either in the laboratory or from a supplier

[EN 12322]

3.4.4

working culture

a primary sub-culture from a reference stock (3.4.3)

4 Practices for quality control of culture media

4.1 Documentation

4.1.1 Documentation required from manufacturer

The following details should be available from the manufacturer:

- name of the medium, individual components and any supplements, and their product codes;
- batch code;

- pH value of the medium before use;
- storage information and expiry date;
- any performance evaluation and test organism used;
- technical data sheet;
- quality-control certificate;
- safety and/or hazard data where needed.

4.1.2 Check list by the laboratory

Laboratory checks upon receipt of the medium:

- name of medium and batch code :
- date of receipt;
- expiry date;
- condition and integrity of packaging;

4.2 Storage

4.2.1 General

In all cases follow the manufacturer's instructions where available regarding storage conditions, expiry date and use.

4.2.2 Quality management and control for dehydrated media and supplements

Media are usually purchased from commercial manufacturers. They are delivered in dehydrated powdered or granulated form in sealed containers and supplements of different selective or diagnostic substances are supplied in either the lyophilised or liquid state. However, purchases should be planned to encourage a regular turnover of stock (i.e. first in-first out). To maintain an effective inventory further checks should include:

- re-checking of the seal;
- date of first opening;
- visual assessment of contents of opened containers.

Especially after opening a new container, the quality of the medium may depend on the storage environment. Loss of quality of dehydrated media is shown by change in flow characteristics of the powder, homogeneity, caking, colour changes etc.. Any dehydrated medium that has absorbed moisture or shows obvious changes in physical appearance should be discarded.

4.2.3 Commercially supplied ready-to-use media

Follow the manufacturer's instructions regarding storage conditions, expiry date and use.

4.2.4 Media prepared from commercially available dehydrated formulations and basic individual components

The shelf-life of these types of media varies. It is therefore difficult to state general time limits for storage of prepared media. Specific International or National Standards may stipulate specific conditions and shelf life.

Sterilised culture media dispensed in plates, tubes or bottles and reagents which are not used immediately shall be protected against light and desiccation.

Unless a validated expiry date has been established or is specified in the International Standard in question, sterile partially complete media, i.e. media to which final components are added immediately before use, shall be kept in a refrigerator for not more than 3 months or at room temperature for not more than 1 month under conditions which prevent their composition from being modified. However, it is recommended that media to which labile selective supplements have been added should be used on the day of preparation. Solid media containing chemically reactive and / or labile substances should not be stored in bulk for remelting.

Observe any colour change, sign of evaporation/dehydration or microbial growth. Batches of media showing such changes should not be used.

Prior to use or before further heating, it is recommended that the culture media be equilibrated to ambient temperature.

4.3 Laboratory preparation of media

4.3.1 General

The accurate preparation of culture media is one of the fundamental steps in microbiological examination and it shall be given special care.

Respect good laboratory practice and the manufacturer's instructions regarding the handling of dehydrated media and other components, particularly those containing hazardous materials i.e. bile salts or other selective agents.

Where media are prepared from dehydrated commercial formulations follow the manufacturer's instructions precisely. Document all relevant data, i.e. weights/volume, pH, date of preparation, sterilisation conditions, operator.

For media prepared from individual components, follow the recipe precisely and record all details as in (4.1.2) and, in addition, the full identity (i.e. code and batch number) of all the components used.

4.3.2 Water

The water used shall be distilled water or water of equivalent quality i.e. free from substances likely to inhibit or influence the growth of microorganisms under the test conditions. If the distilled water is prepared from chlorinated water, neutralise the chlorine prior to distillation.

The distilled water shall be stored in containers manufactured preferably from inert materials (e.g. neutral glass, polyethylene etc.) which shall be shown to be free of any inhibitory substances prior to their initial use.

NOTE In some cases, it may be necessary to use freshly prepared water, free of dissolved carbon dioxide.

In order to be considered as being of good quality, the distilled water shall have a resistivity of at least $300~000\Omega cm$.

WARNING Water processed through an ion exchanger (de-ionised), may have a high microorganism content; it is therefore advisable not to use such water without verifying that the microorganism content of the water is low. Consult the manufacturer for the best way to minimise microbial contamination. Heavily

contaminated deionised water that has been filter sterilised may still contain substances inhibitory to the growth of some microorganisms.

4.3.3 Weighing and rehydration

Carefully weigh the appropriate amount of dehydrated medium (taking care not to inhale powder, especially with media containing toxic substances) and progressively add the required amount of water avoiding clumping.

4.3.4 Dissolution and dispersion

Dehydrated media needs rapid dispersion by instant and repeated stirring followed by heating, if necessary, to dissolve. Media containing agar should be allowed to soak for several minutes prior to heating with mixing to dissolve. For media prepared from individual components each component should be added separately and allowed to dissolve before finally making up to volume.

4.3.5 Measurement and adjustment of pH

Measure the pH using a pH meter and adjust if necessary i.e. for media prepared from individual components in the laboratory so that after sterilising and cooling to 25 °C the medium is at the required pH \pm 0,2 pH units, unless otherwise stated. The adjustment is normally carried out using a solution of approximately 40g/l (about 1 mol/l) of sodium hydroxide (NaOH) or approximately 36,5 g/l (about 1 mol/l) hydrochloric acid (HCl).

NOTE Commercially manufactured media may show significant changes in pH before and after autoclaving. However, provided good quality distilled or de-ionised water is used, pH adjustment prior to autoclaving should not be necessary.

4.3.6 Dispensing

Dispense the medium into appropriate containers having a volume 1,2 to 3 times that of the medium.

4.3.7 Sterilisation

4.3.7.1 General

The sterilisation of culture media and of reagents may be carried out by using sterilisation by moist heat (4.3.7.2) or sterilisation by filtration (4.3.7.3).

Certain media do not need sterilisation by autoclaving but can be used following boiling. For example, media for *Enterobacteriaceae* containing brilliant green are particularly sensitive to heat and light and should be rapidly cooled after boiling and protected from strong light. Also some reagents can be used without sterilisation (refer to appropriate International standard or manufacturer's instructions).

4.3.7.2 Sterilisation by moist heat

Sterilisation by moist heat is performed in an autoclave or media preparator. Generally the autoclaving operation takes 15 min at 121 °C. For volumes greater than 1 000 ml, adapt the sterilisation cycle as necessary. In all cases follow the instructions given in the International Standard or the manufacturer's instructions. The performance of the autoclave should be monitored by temperature profiling using thermocouples and test strips under typical load conditions to ensure the desired temperatures can be reached.

NOTE Overheating can occur when large volumes of media (> 1 000 ml) are processed in an autoclave.

Control of the efficacy of sterilisation is essential.

After heating it is essential that media be cooled in a manner to prevent boiling over. This is particularly important for sensitive media e.g. *Enterobacteriaceae* and media in large volumes.

4.3.7.3 Sterilisation by filtration

Sterilisation by filtration can be performed under vacuum or pressurised conditions. Use membranes and filter elements with a pore diameter of $0.22~\mu m$. They shall have been sterilised in the autoclave. Refer to the manufacturer's instructions regarding the use of filter elements or membranes that have been purchased in a sterile condition. Sterilise the different parts of the filtration apparatus, assembled or not, in the autoclave for 15 min at $121~^{\circ}C$. If necessary, aseptic assembly can be performed in a laminar flow cabinet after autoclaving.

NOTE Some filter membranes may retain proteins (such as antibiotics). In order to obtain the correct concentration the user should pre-wet the filter.

4.3.7.4 Monitoring

After autoclaving, boiling or filtration, all media should be monitored, in particular with respect to pH, colour, sterility and consistency.

4.3.8 Preparation of supplements

Manufactured supplements containing toxic agents, particularly antibiotics, must be handled with care avoiding dispersion of powder which may give rise to allergic or other reactions in laboratory personnel. Take appropriate precautions and follow the manufacturer's instruction when making solutions. Do not use beyond their stated shelf-life which, for antibiotic working solutions, is generally the same day. Under certain circumstances, antibiotic solutions may be stored frozen in suitable aliquots but should not be re-frozen after thawing. The potential loss of activity due to freezing should be discussed with the manufacturer or tested by the user.

4.4 Preparation for use

4.4.1 Melting of agar culture media

Melt a culture medium by placing it in a boiling water bath or by any other process which gives identical results (e.g. a steam flow-through autoclave). Media that have previously been autoclaved should be reheated for a minimum time to maintain media quality. Avoid over-heating and remove when it has melted. Cool the molten medium to 47 $^{\circ}$ C \pm 2 $^{\circ}$ C in a thermostatically controlled water bath until such time as it is to be used. The time needed to reach 47 $^{\circ}$ C depends on the type of medium, the volume and the number of units in the water bath. Molten medium should be used as soon as possible but it is recommended that it should not be retained for more than 4 h.

4.4.2 De-aeration of culture media

If necessary, just prior to use, heat the culture medium in boiling water or under a flow of steam for 15 min, with lids or caps loose; after heating, tighten the caps and cool down rapidly to the operating temperature.

4.4.3 Addition of supplements

Heat-labile supplements should be added to the medium after it has been cooled to $47^{\circ}C \pm 2^{\circ}C$. Allow the sterile supplement to come to room temperature before adding it to the agar medium. Cold liquids may cause agar to gel or form transparent flakes. Mix all supplements into the medium gently and thoroughly, then distribute into the final containers as quickly as possible.

4.4.4 Preparation and storage of media in Petri dishes

Pour the molten agar culture medium into Petri dishes so as to obtain a thickness of at least 2 mm (e.g. for 90 mm diameter dishes, 15 ml of agar are normally required). Allow the agar to cool and solidify by placing the Petri dishes with lids in place on a cool, horizontal surface.

NOTE During incubation, a loss of moisture of the agar media will occur. A loss of more than 15 % of the water content can adversely affect the growth of microorganisms in some circumstances. Factors influencing water loss are medium composition, amount of medium in the plates, the type of incubator i.e. fan-assisted or otherwise, humidity of the atmosphere in the incubator, the position and number of the plates in the incubator and the incubation temperature.

Use the solidified medium immediately or store under conditions which prevent its composition from being modified, i.e. in the dark and/or in the refrigerator at 4°C to 12°C in sealed bags for a maximum period of one week or as directed by the manufacturer or specific standard. Label the dishes on the base with date of preparation and/or expiry date and identity. Alternative coding systems meeting these requirements may be used.

The shelf-life of poured plates will increase if they are stored in sealed plastic bags. In order to avoid the occurrence of condensate, the plates must be cool before being placed into bags. Do not dry the surface of agar plates prior to chill storage.

In general, for the surface inoculation of a solid culture medium, dry the dishes, preferably with the lids removed and with the agar surface facing downwards, in an oven set at a temperature between 25 °C and 50 °C or in a laminar-flow cabinet, until the droplets have disappeared from the surface of the medium. Do not over-dry them. Commercially prepared ready-to-use agar plates should be stored and used according to the manufacturer's instructions.

4.4.5 Incubation

Do not stack dishes in piles more than 6 high and leave space for air circulation to allow medium to equilibrate to incubation temperature as rapidly as possible. For liquid media, time to reach incubation temperature is dependant upon a number of factors e.g. volume, loading, container, incubator type. In the case of anaerobic jars it may be necessary to stack plates in excess of 6 high.

4.5 Disposal of media

Both contaminated and unused media must be disposed of in a manner that is safe and meets any local or national Regulations.

5 Quality control of finished product

5.1 Physical quality control

Laboratory testing should include as a minimum:

pH-value measured at between 20 °C and 25 °C;

and by observation:

- quantity filled and/or layer thickness;
- colour;
- clarity/presence of optical artefacts;
- gel stability / consistency / moistness.

5.2 Microbiological quality control

5.2.1 Contamination

An appropriate amount of each batch should tested for contamination.

5.2.2 Test organisms

A set of test organisms should only contain microorganisms with stable characteristics representative of their species and that have been shown to be reliable for the demonstration of optimal performance of a particular laboratory prepared medium. The test organisms should primarily comprise strains that are widely available in reference culture collections, but well-characterised strains isolated by the laboratory may also be included. The relevant cultural characteristics of the stock culture should be examined and recorded by the laboratory and the strain renewed should atypical characteristics occur. It is preferable to use strains that have originated from foods although not all culture collections provide such data on their origin.

The test organisms for each medium may include:

- robust positive strains with typical characteristics;
- weakly growing positive strains (i.e. of a more sensitive nature);
- biochemically un-reactive strains e.g. those showing different fermentation or fluorescence reactions;
- completely inhibited strains.

NOTE The International Committee for Food Microbiology and Hygiene's Working Party on Culture Media (WPCM) have prescribed a validated collection of test strains for media evaluation [1].

5.2.3 Ready-to-use media and reagents

Manufacturers of commercially available ready-to-use media especially if approved to ISO 9001 [2] or ISO 9002 [3] standards, will have a quality programme in place and may issue a quality certificate with the media they supply. Under those conditions the user may not need to carry out extensive testing on such media but should ensure that storage conditions are maintained.

5.2.4 Media prepared from commercially available dehydrated formulations

For the purposes of this part of ENV ISO/TR 11133-1, qualitative tests for each batch of prepared medium are the minimum requirements but, where quantitative examinations on samples are to be carried out, quantitative tests on each batch will give greater assurance of media quality. For those media which contain no indicators or selective agents the use of a single positive test strain is appropriate. For those media which do contain indicators or selective agents, strains which demonstrate the function of the indicator(s) and selectivity must be utilised. For complex media, i.e. with added supplements, each batch should be verified with strains with characteristics listed in 5.2.2. In the case of ready-to-use media to which laboratory-prepared supplements have been added the same applies.

5.2.5 Media prepared from basic individual components.

It is recommended that in addition to the qualitative tests described in 5.2.4 that some quantitative testing is carried out using techniques such as the modified Miles & Misra technique [1] or spiral plating in order to monitor trends in quality of basic materials, productivity of the medium and in-house laboratory production protocols.

NOTE These are minimal guidelines, in practice, foods may contain stressed microorganisms. The suitability of the medium with respect to the recovery of stressed cells should be taken into account. For information on preservation, maintenance techniques and reference culture collections see Annex B.

Annex A

(informative)

Designation of the components of the culture media in standards on microbiological analysis of food and animal feeding stuffs products

A.1 General

In order to harmonise the description of the various components in the composition of culture media in microbiological standard methods, ISO/TC 34/SC 9 "Food and agricultural products —Microbiology" decided the designations for the categories of components designated in A.2 to A.5.

A.2 Peptones

-	enzymatic digest of casein ²⁾ ;
-	enzymatic digest of soybean meal;
-	enzymatic digest of animal tissues ³⁾ ;
-	enzymatic digest of heart;
-	enzymatic digest of gelatin;
-	enzymatic digest of animal and plant tissue.
A. 3	BExtracts
A.3	BExtracts meat extract;
A.3	
A.3	meat extract;
A.3	meat extract ; brain-heart extract ;

A.4 Agar

bacteriological agar;

bile salts No.3.

A.5 Other

-	egg yolk emulsion;

²⁾ This includes peptic digest of casein, tryptic digest of casein and tryptone.

³⁾ This includes meat peptone, peptic digest of meat, pancreatic digest of meat.

- skim milk powder;
- acid hydrolysate of casein.

Annex B

(informative)

Guidance on preservation and maintenance of control strains

B.1 General

There are several methods available, i.e. lyophilisation, storage on beads at -70 °C, or using liquid nitrogen, for the successful preservation and maintenance of all microorganisms relevant to food microbiology. One method may not be appropriate for all strains.

These guidelines are presented in figure B.1.

B.2Reference strains from commercial sources

If reference strains from reference collections or ISO 9001 [2] or ISO 9002 [3] approved commercial suppliers are purchased and maintained in their original containers, the manufacturers directions for their cultivation and use must be followed.

B.3Laboratory-prepared reference stocks

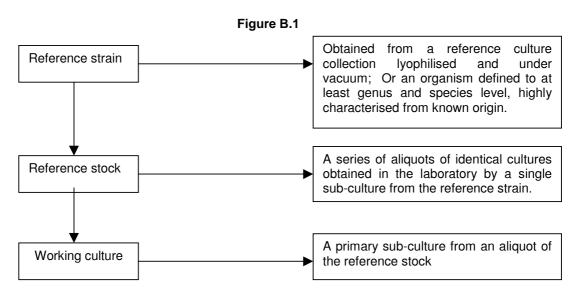
Stock cultures of reference strains (B.2) for quality control and performance testing purposes must be maintained and handled in a manner, which minimises the opportunity for cross-contamination, mutation or alteration of typical characteristics. Reference stocks should be stored in multiple aliquots either deep-frozen (-70°C) or lyophilised. Their growth characteristics should be fully documented for each medium on/in which they will be utilised as test organisms.

B.4Working cultures

Working cultures are prepared from lyophilised or deep-frozen reference stocks (B.3). Aliquots must be handled in a manner, which avoids possible cross-contamination of the reference stock and/or its deterioration. Working cultures should be prepared by resuspending an aliquot of the reference stock in a non-selective growth medium and incubating to yield a stationary phase culture.

For commercially available preservation systems the manufacturer's instructions shall be rigorously followed.

NOTE Sub-cultures from working cultures should not be made. However, working cultures may be used more than once provided they are handled and stored appropriately i.e. to avoid cross-contamination and/or deterioration for no longer than one week.



Annex C (informative) Quality assurance of culture media – trouble shooting

Abnormality	Possible reason
Agar medium fails to solidify	Overheating of medium during preparation Low pH causing acid hydrolysis to occur Incorrect weight of agar used Agar not thoroughly dissolved Poor mixing of ingredients
Incorrect pH	Overheating of medium during preparation Poor water quality Extraneous chemical contamination pH measured at incorrect temperature pH meter incorrectly calibrated Poor quality dehydrated medium
Abnormal colour	Overheating of medium during preparation Poor water quality Poor quality dehydrated medium Incorrect pH Extraneous contamination
Formation of precipitates	Overheating of medium during preparation Poor water quality Poor quality dehydrated medium Poor pH control
Medium inhibitory/Low productivity	Overheating of medium during preparation Poor quality dehydrated medium Poor water quality Incorrect formulation used, e.g. ingredients not weighed out correctly, supplements added at wrong concentration
Poor selectivity	Overheating of medium during preparation Poor quality dehydrated medium Incorrect formulation used Supplements added incorrectly e.g. when medium too hot or at wrong concentration

Bibliography

- [1] Culture media for Food Microbiology. Progress in Industrial Microbiology, Vol. 34 (1995). Eds. Corry J.E.L., Curtis, G.D.W and Baird, R.M. Elsevier, Amsterdam.
- [2] ISO 9001, Quality systems —Model for quality assurance in design, development, production, installation and servicing.⁴⁾
- [3] ISO 9002, Quality systems Model for quality assurance in production, installation and servicing. 4)

⁴⁾ These will be combined as ISO 9001:2000, Quality management systems - Requirements.